

Environmental DNA surveys of fish community composition across the mid-Atlantic region: One method, many habitats

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- 6 – NJ DEP

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Applications

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A collage of marine life. In the upper left, a whale is breaching. In the center, a large fish swims towards the viewer. In the upper right, a boat is on the water. In the upper left, several birds are flying. In the lower left, a school of small fish swims. In the lower right, a large fish swims away.

Communities

- The representative assemblage of organisms at a given space / time
- Emergent ecological property
 - Complex systems with many interaction
 - Resilience

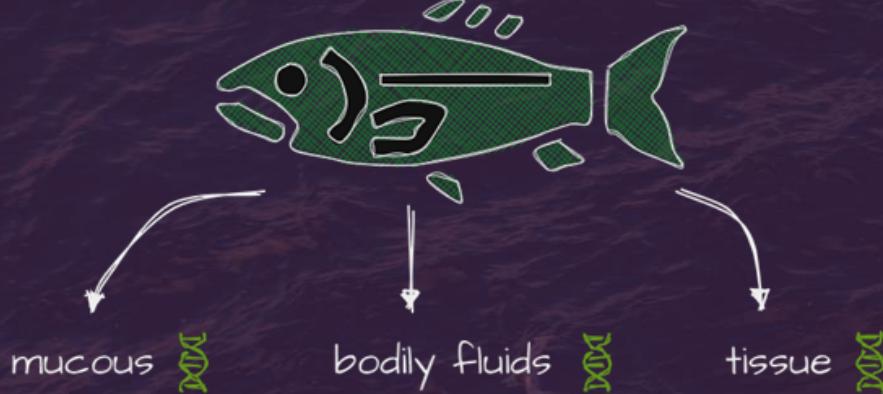
Community composition is a good ecological indicator, but measurement across different habitats can be challenging.

eDNA metabarcoding can help with that.

What is eDNA?

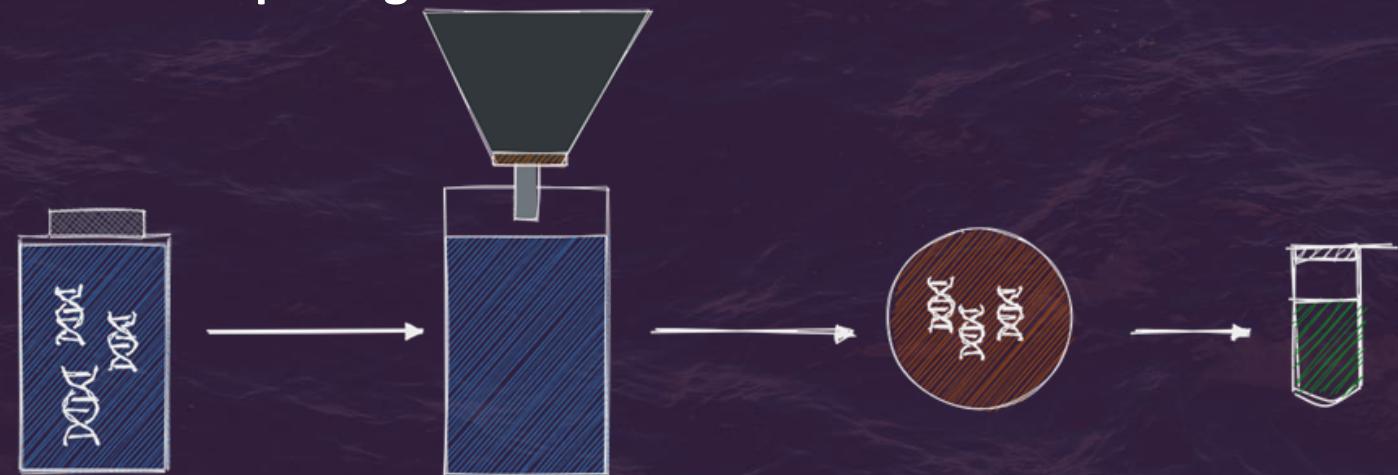
DNA is shed as cellular or extracellular material into the surrounding water

Marine eDNA has ~ 1 d half-life



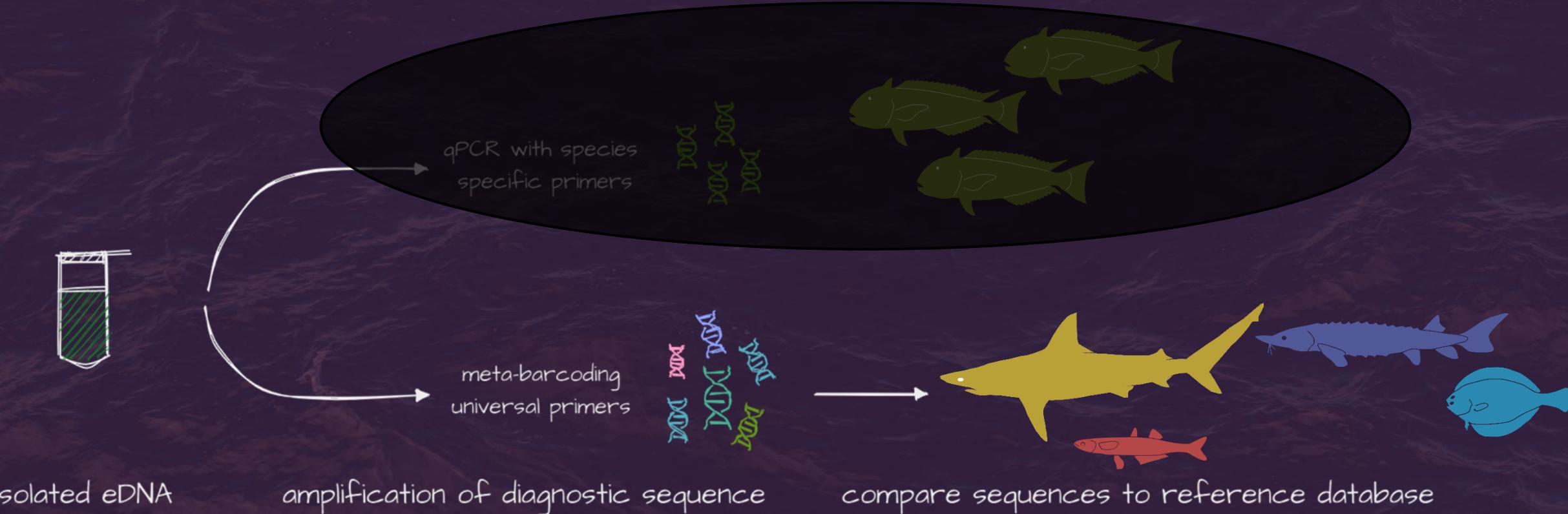
collect & filter water from aquatic systems

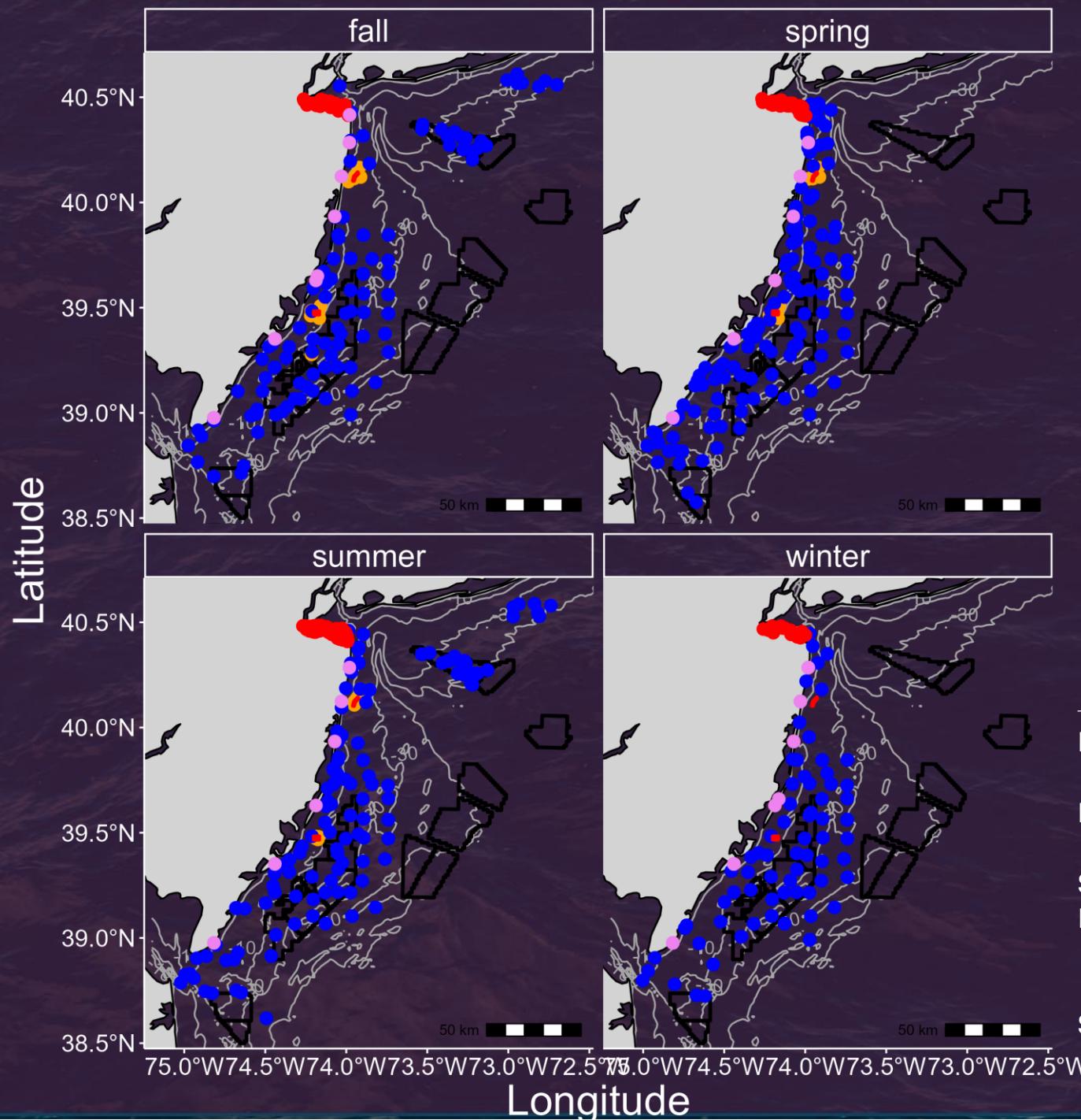
Bottom waters sampled with some surface pairing



Environmental DNA = DNA isolated from an environmental sample

eDNA metabarcoding = robust community data





The Dataset and habitat designations:

736 total samples analyzed

143 amplicon sequence variants identified

Sample sizes by season

habitat	fall	spring	summer	winter
estuary	24	30	94	14
reef	16	14	12	
shelf	129	126	135	107
surf zone	11	6	6	12

Habitats by Campaigns

Estuary – Raritan Inventory Project (RIP)
(Stacy van Morter)

Reef – NJ Artificial Reefs program
(Peter Clarke)

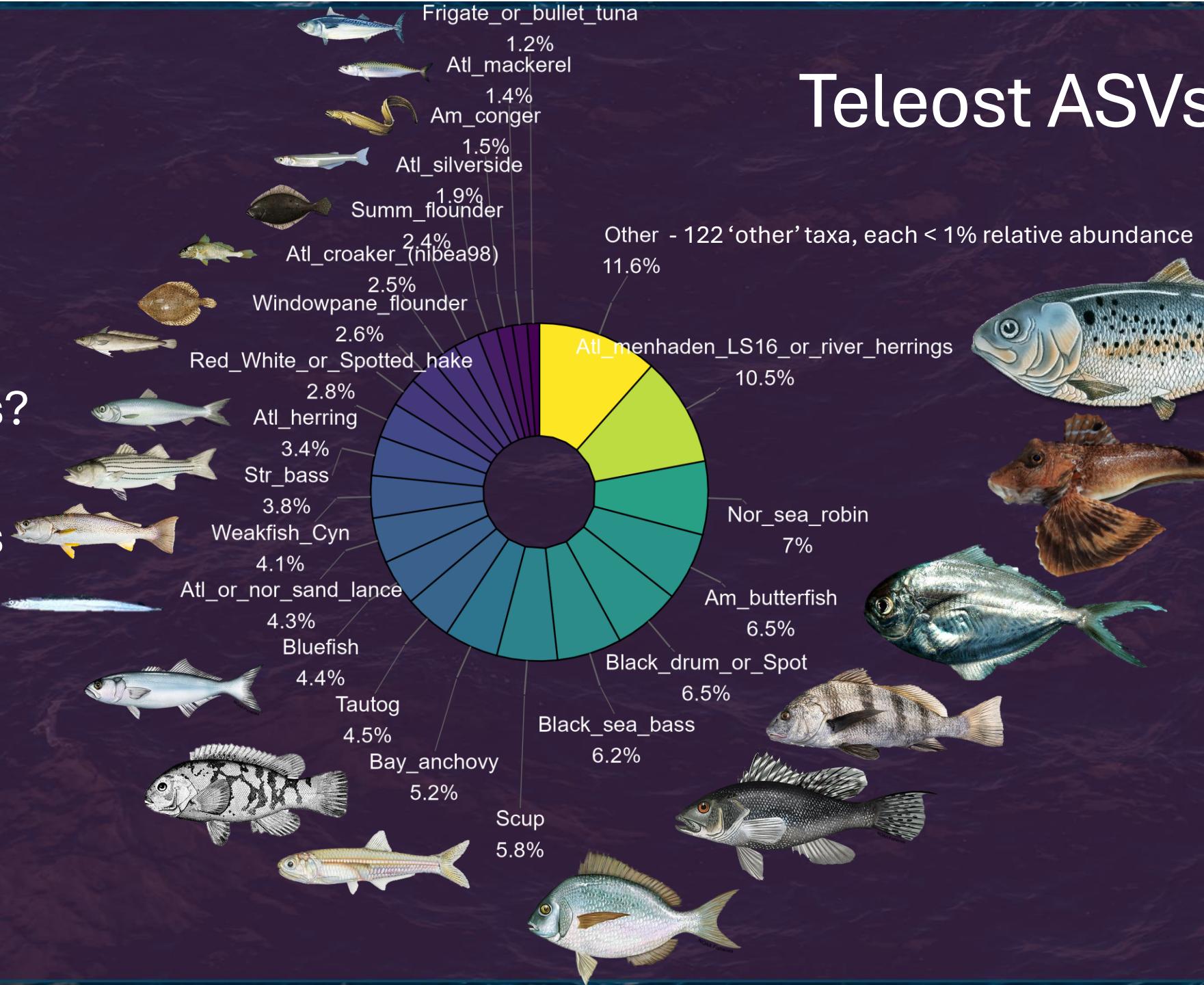
Shelf – NJ Ocean Trawl, Monmouth University 'RMIO', & Inspire Environmental surveys
(Gregory Hinks (DEP); Brian Gervelis and Dara Wilber (Inspire Environmental))

Surf zone – Monmouth University and community scientists

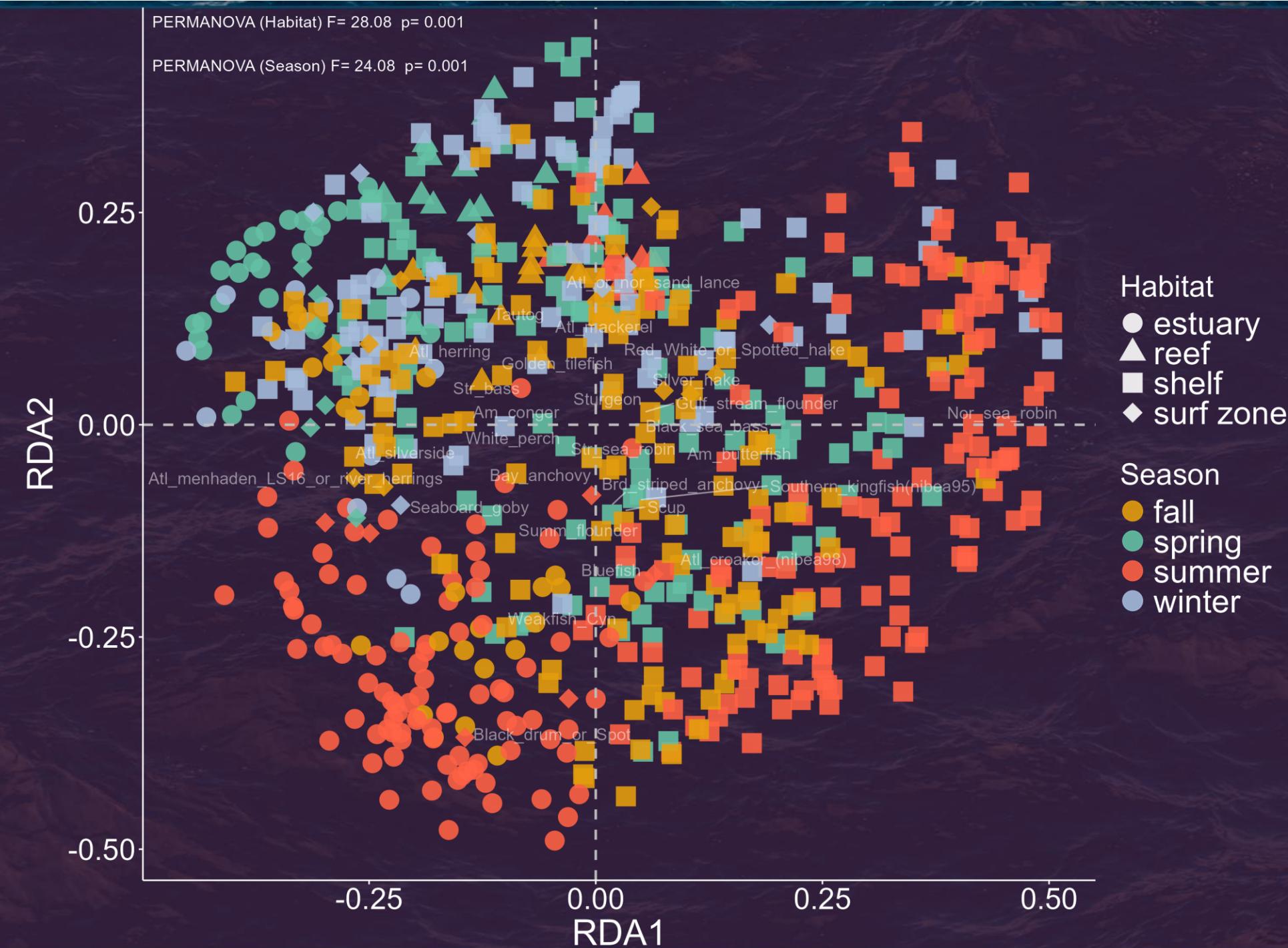
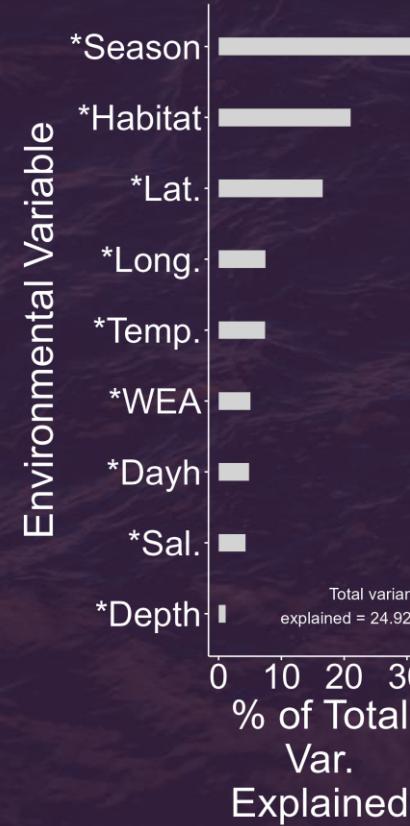


Teleost ASVs

- How are they organized in communities?
- How are communities related to habitat?

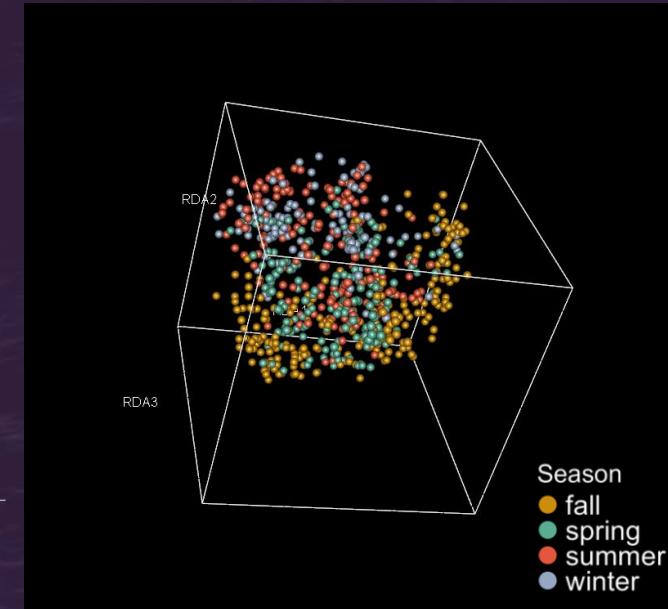
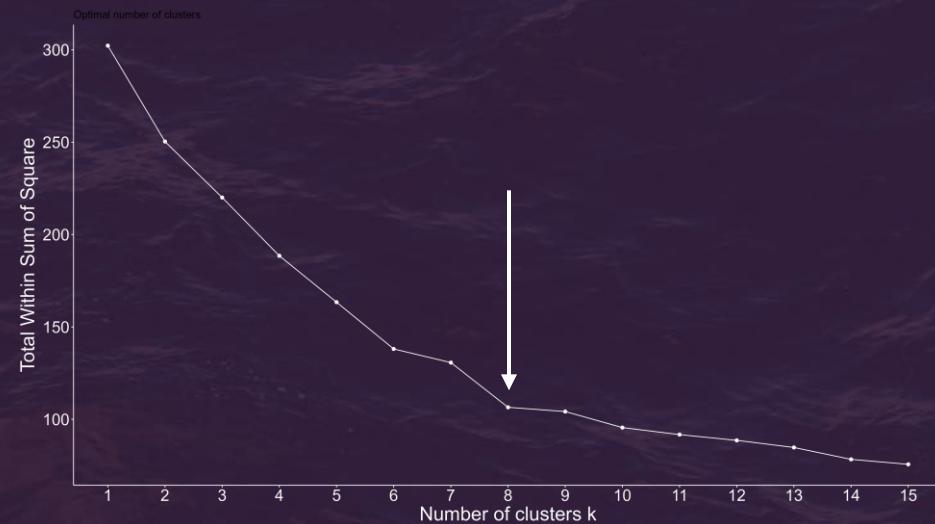


Ordination of community data



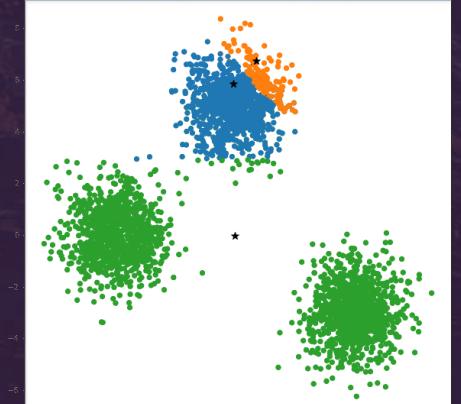
K-means clustering to label communities

- Cluster results for RDA scores
 - Clusters in 6D data space from RDA scores
 - How many ‘named’ communities are there and how well does clustering work?
 - Elbow plot for choosing k-clusters: **8** clusters
 - Avg silhouette width: **0.28** (‘meh’)
 - PERMANOVA % variance in original fish community matrix explained – **35%**



What our data actually looks like, in 3 out of 6 dimensions after RDA

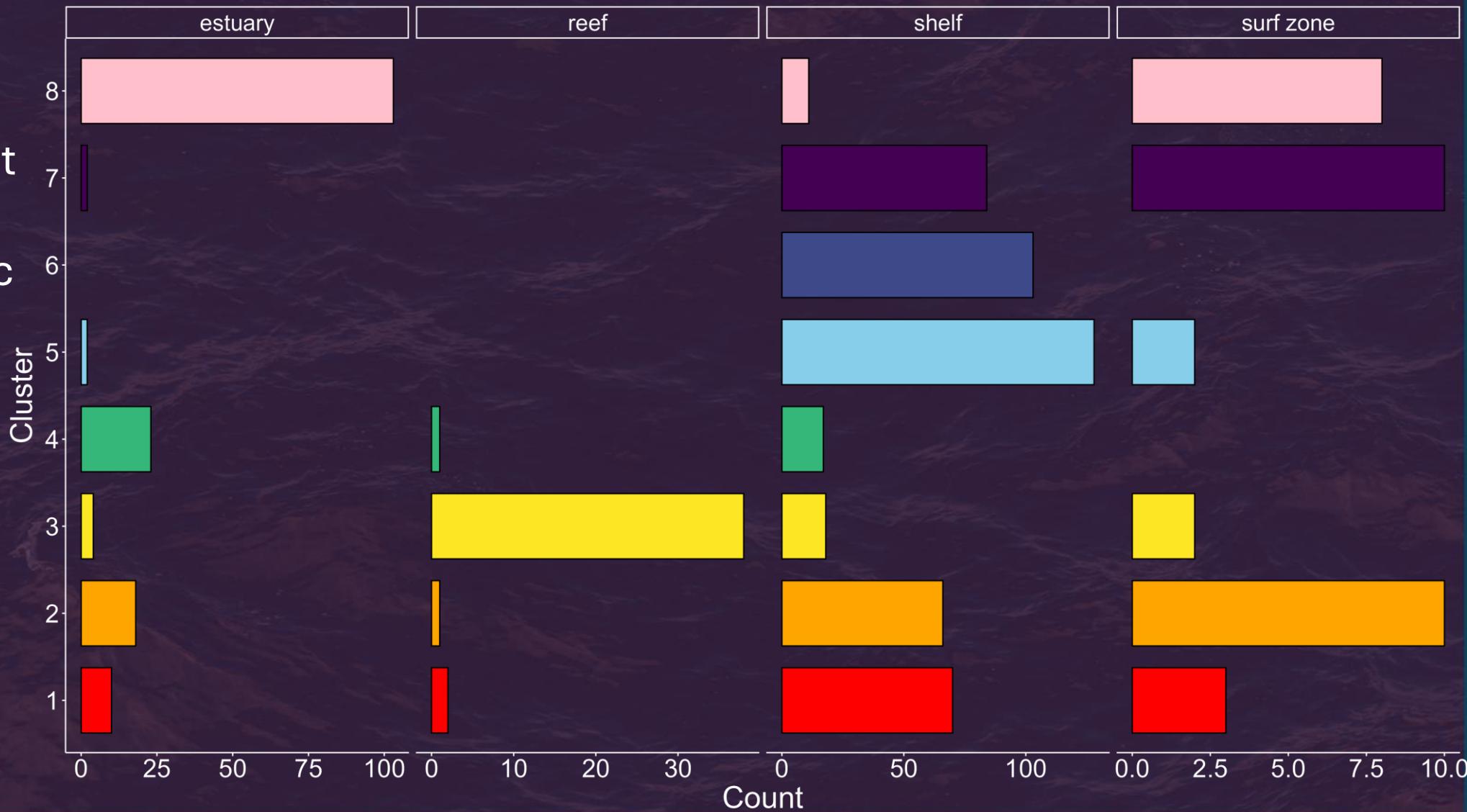
These data are complex beyond 8 clusters, but this is still a useful approach to understanding the data better



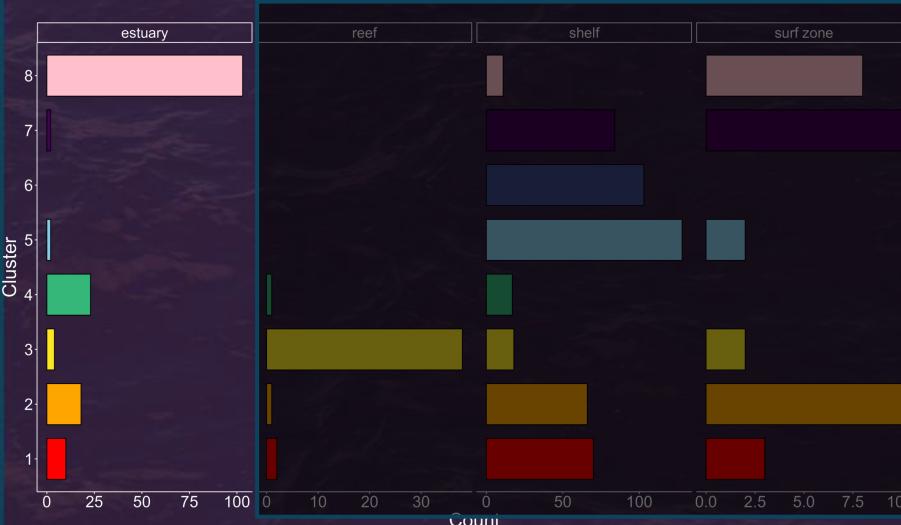
<https://towardsdatascience.com/unsupervised-learning-k-means-clustering-6fd72393573c/>

Certain clusters were dominant in certain habitats

This indicates that habitats sampled had characteristic fish community composition as determined by eDNA metabarcoding



ESTUARY



You're turn to guess:

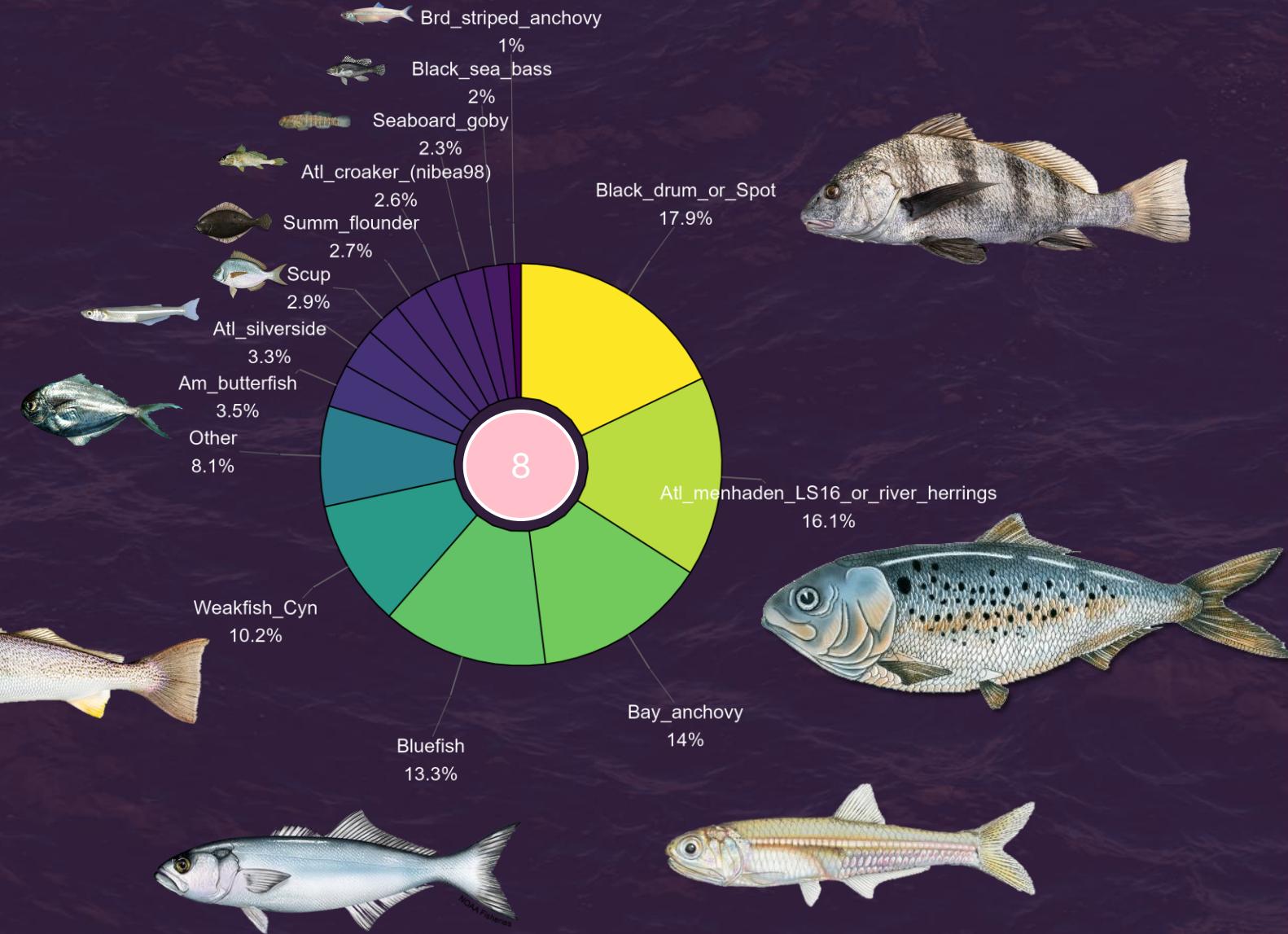
SURF ZONE?

SHELF?

ARTIFICIAL REEF?

Or

ESTUARY?



REEF



You're turn to guess:

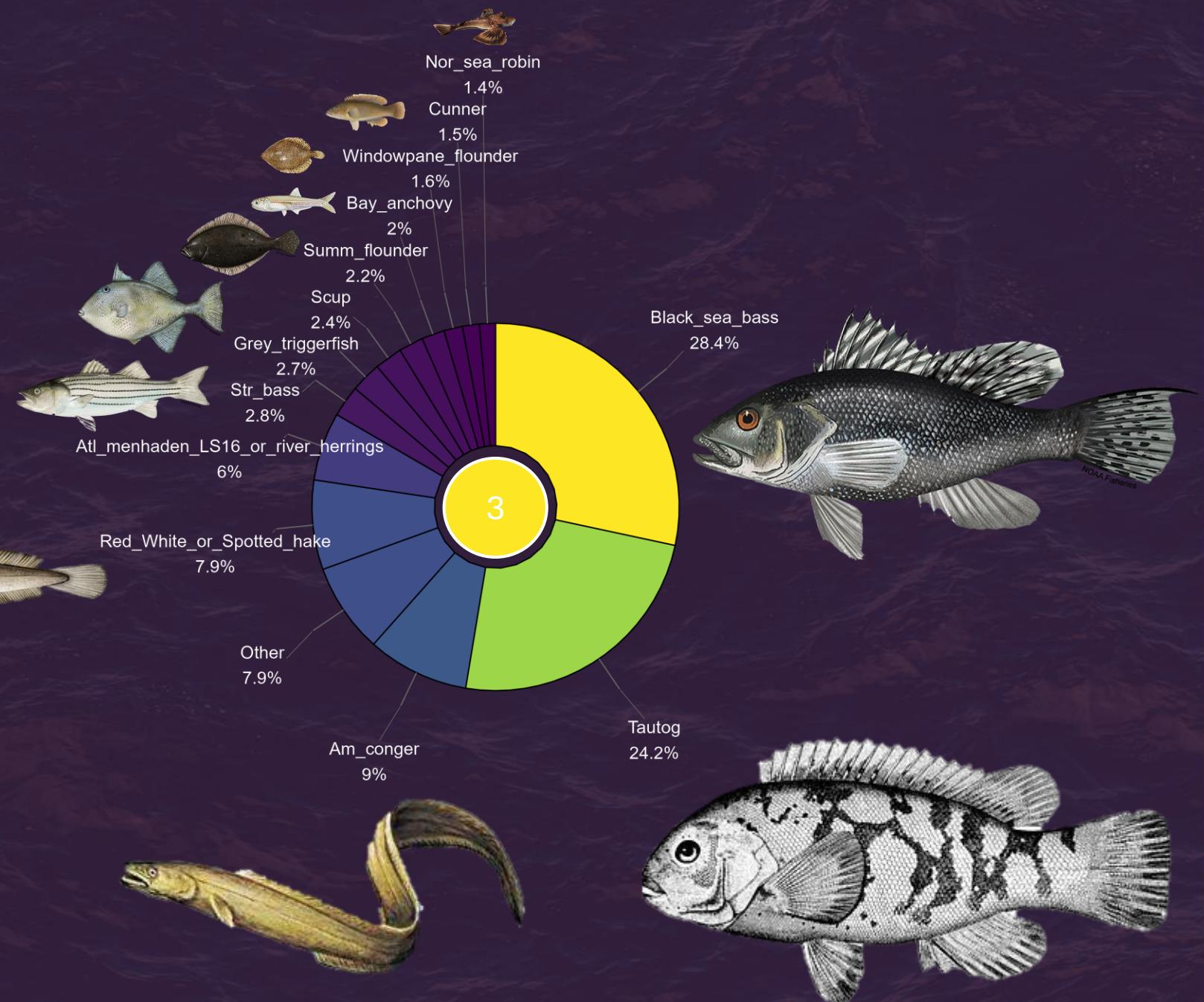
SURF ZONE?

SHELF?

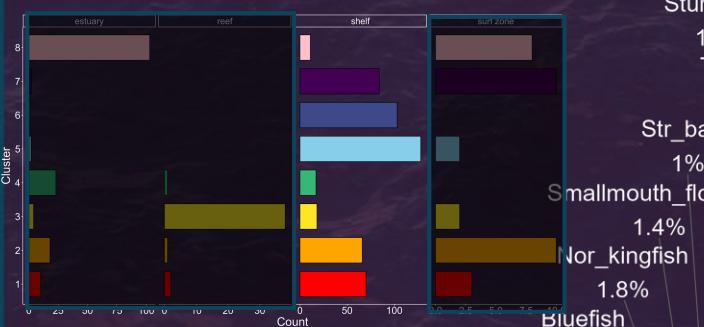
ARTIFICIAL REEF?

Or

ESTUARY?



SHELF



Sturgeon

1%

Tautog

1%

Str_bass

1%

Smallmouth_flounder

1.4%

Nor_kingfish

1.8%

Bluefish

1.8%

Am_butterfish

11.1%

Red_White_or_Spotted_hake

2.8%

Black_sea_bass

3.3%

Bay_anchovy

3.3%

Brd_striped_anchovy

3.3%

Scup

4.3%

Southern_kingfish(nibe95)

5%

Atl_menhaden_LS16_or_river_herrings

5.3%

Atl_croaker_(nibe98)

5.3%

Summ_flounder

5.7%

Windowpane_flounder

6.2%

Weakfish_Cyn

6.3%

Nor_sea_robin

8.4%

Black_drum_or_Spot

11%

Am_butterfish

5.1%

Bluefish

7.4%

Other

10.8%

Nor_sea_robin

8.4%

Weakfish_Cyn

6.3%

Summ_flounder

5.7%

Windowpane_flounder

6.2%

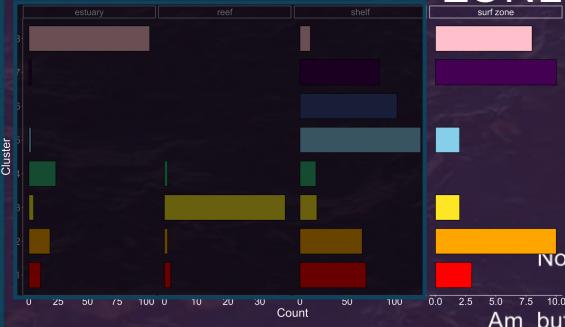
Nor_sea_robin

8.4%

Weakfish_Cyn

6.3%

SURF ZONE



You're turn to guess:

SURF ZONE?
SHELF?
ARTIFICIAL REEF?
Or
ESTUARY?



Black_sea_bass
1%
Black_drum_or_Spot
1.6%
Nor_sea_robin
1.6%
Atl_herring
1.6%
Nor_pipefish
1.8%
Am_butterfish
2.3%
Str_bass
2.3%
Atl_croaker_(nibe98)
2.3%
Smallmouth_flounder
2.4%
Summ_flounder
2.7%
Red_White_or_Spotted_hake
3.6%
Bay_anchovy
4.2%
Atl_silverside
5.3%
Atl_mackerel
8.3%
Windowpane_flounder
9%
Other
9.8%



Atl_menhaden_LS16_or_river_herrings
40.1%



Smallmouth_flounder
2.4%



Summ_flounder
2.7%



Red_White_or_Spotted_hake
3.6%



Bay_anchovy
4.2%



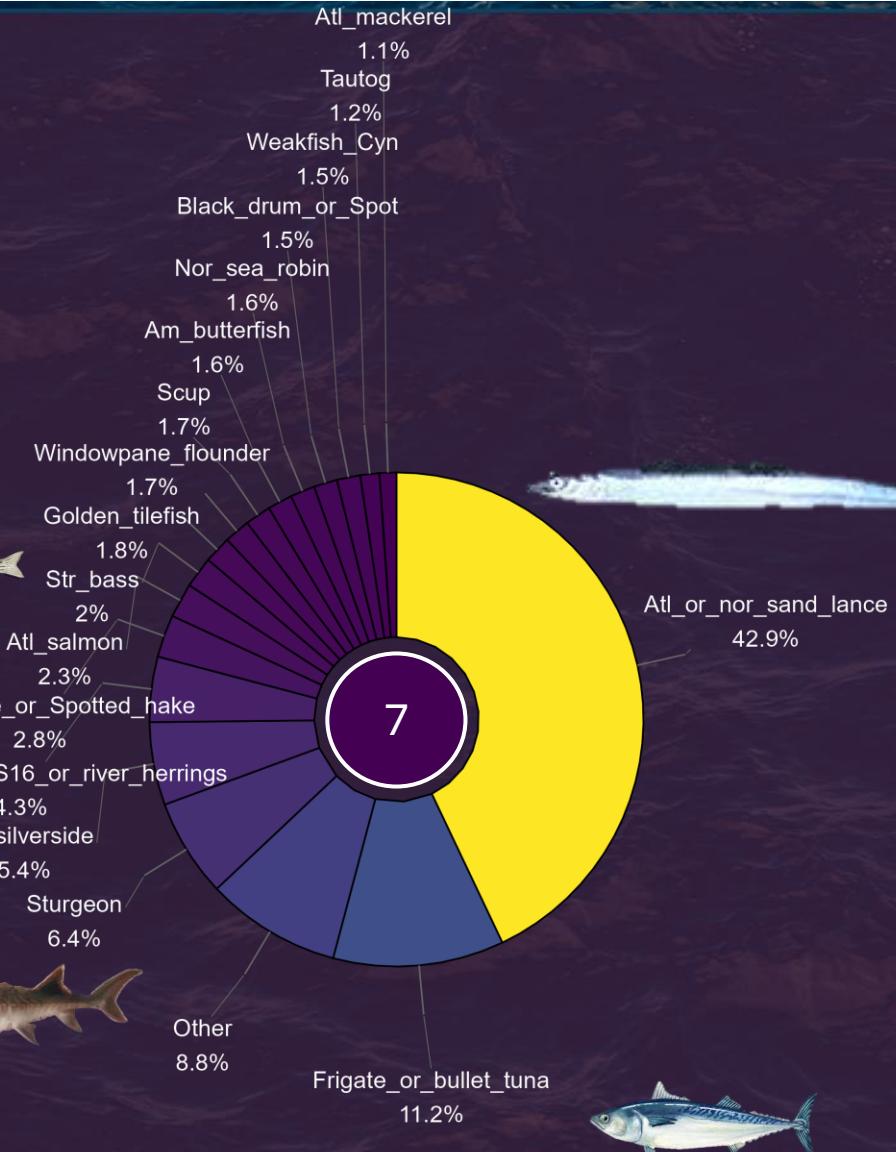
Atl_silverside
5.3%



Atl_mackerel
8.3%

2

2



Atl_or_nor_sand_lance
42.9%



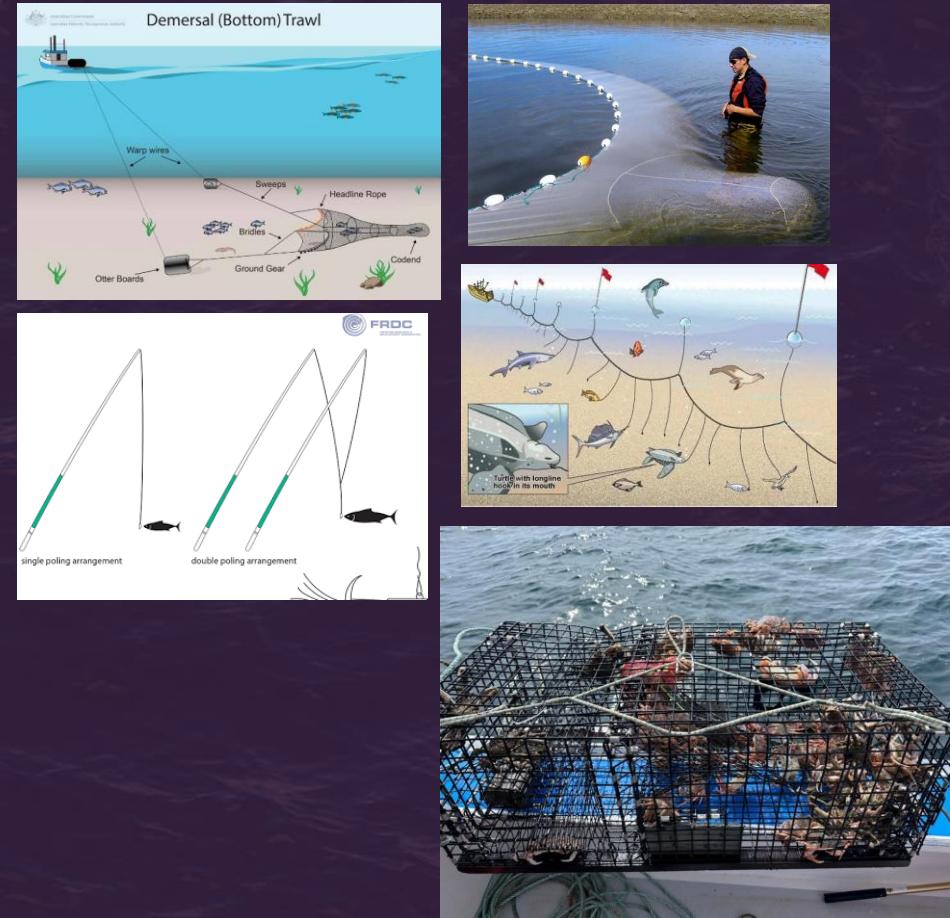
3

3

3

What's it look like if we do this analysis on capture data?

- You can't... that's *kinda* the point
- eDNA allows sampling across habitats and taxa with consistent bias
 - Clark et al. - River Center Conference Rm 18, 10:30 AM (TODAY!)
 - There's more than fish that may be useful in 12S metabarcoding data
 - O'Leary et al. River Center Conference Rm 18, 9:30 AM (TODAY! NEXT!)



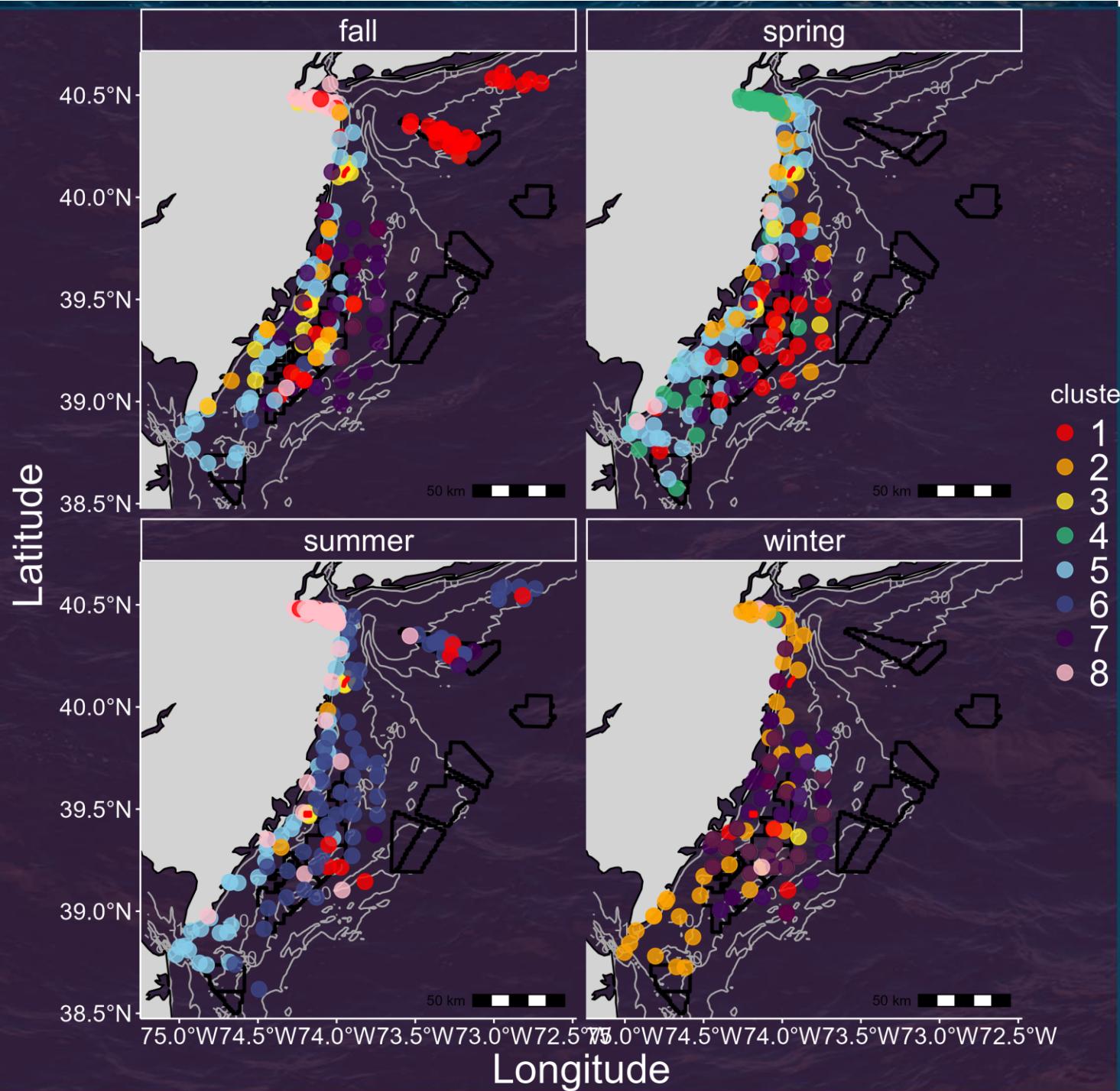
Why is this useful?

Community composition is a good and resilient ecological indicator, but can be complex, hard to measure and difficult to visualize / summarize

We have far better methods for analyzing community data than for collecting it broadly

eDNA alleviates the collection issue, allowing us to focus more on the analyses!

- Leaflet interactive map



Conclusion

- eDNA metabarcoding provides robust community composition data that is a logical ‘response variable’ for impact-type studies
 - Community composition is a robust index based on a lot of data that integrates biological – biological and biological – physical interactions
 - Offshore wind development BACI / BAG
 - Restoration
 - Oyster reefs
 - Seagrasses
 - Climate research
 - ‘events’ causing community shifts



Equinor is beginning installation of turbines at the Empire wind site.



Funding:



Colleen Brust & Caitlin McGarigal (NJDEP Project Managers)



NJ DEP capture survey contacts

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NJAR – Peter Clark

NJRIP – Stacey van Morten



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